# CURRENT CONCEPT REVIEW

# Avascular Necrosis of the Femoral Head: Are Any Genes Involved?

Farzaneh Pouya, MSc; Mohammad Amin Kerachian, MD, PhD

Research performed at Medical Genetics Center, Mashhad University of Medical Sciences, Mashhad, Iran

Received: 17 December 2014

Accepted: 4 May 2015

# Abstract

Avascular necrosis of the femoral head (ANFH) is a pathologic process that results from interruption of blood supply to the femur bone resulting in the death of bone cells and collapse of the femoral head. Nontraumatic ANFH continues to be a significant challenge to orthopedic surgeons. While the exact mechanisms remain elusive, many new insights have emerged from research in the last decade that has given us a clearer picture of the pathogenesis of nontraumatic ANFH. Progression to the end stage of ANFH appears to be related to five main mechanisms: hypercoagulable conditions, angiogenesis suppressions, hyperadipogenesis, heritable states, and switching the bone remodelling into bone resorption. Researchers have been examining the pathogenic mechanisms of ANFH but none of these theories have been firmly confirmed although some appear more plausible than the others. All of these factors can switch bone remodelling into bone resorption, which can further lead to ANFH progression ending up to femoral head collapse.

Key words: Avascular, Avascular necrosis, Femur, Osteonecrosis

## Introduction

A vascular necrosis of the femoral head (ANFH) is manifested with death of bone cells resulting in the impairment of normal reparative processes within the microfractures in the femoral head (1). Because the exact pathophysiology is not elucidated yet, there is a variable nomenclature for this condition. The name avascular necrosis (also known as osteonecrosis, aseptic, or ischemic necrosis) may be misleading as it has not been demonstrated that bone cells die because of necrosis. In the other words, characteristics of necrosis in the soft tissue including cell swelling and inflammatory responses do not occur in ANFH (2).

inflammatory responses do not occur in ANFH (2). Its true prevalence of the disease is unknown. Approximately 10,000 to 20,000 new patients with ANFH are diagnosed each year in the United States and there are 300,000-600,000 people having the disease (3). Almost 75% of patients with ANFH are between 30 to 60 years of age (4). Since most patients with the disease are at the peak of their productive years, there is a considerable effect on the workforce and subsequently on the economics. The understanding

**Corresponding Author:** Mohammad Amin Kerachian, Medical Genetics Research Center, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran E-mail: Kerachianma@mums.ac.ir

of its pathophysiology and its progression will help us prevent patients, their family and society from these consequences.

A number of studies have examined the pathogenic mechanisms of avascular necrosis in the femoral head and none of the current hypotheses have been firmly established although some appear more plausible than others. In this review, different molecular mechanisms and several mediators important in pathogenesis of ANFH are going to be discussed.

#### **Coronary disruption of the hip**

# Role of Endothelium in Avascular Necrosis

Although the pathophysiology of non-traumatic ANFH remains unclear, most scholars agree that ANFH is associated with thrombus formation in the microvasculature of the femoral head as depicted in the vascular hypothesis for ANFH (1). It appears that damage to the endothelial cell membrane combined in some cases with an increased propensity to blood clot formation, is the key causing blood flow interruption. The endothelial cell monolayer constitutes the inner



THE ONLINE VERSION OF THIS ARTICLE ABJS.MUMS.AC.IR

Arch Bone Jt Surg. 2015;3(3):149-155.

lining of the vascular wall and plays an essential role in the homeostasis of the blood vessel. Due to its unique localization, the endothelium is continuously exposed to inflammatory cells and circulating factors, which could induce endothelial activation and/or endothelial injury (5). In a study by Jacobs et al. damage or abnormality of the reticular vessels was suggested to be the underlying mechanism of ANFH (6). Damage to the endothelial cells may result in abnormal blood coagulation and thrombi formation. Thus, avascular necrosis could occur distal to the site of arterial occlusion. Slichter et al. postulated that endothelial cell damage is followed by platelet thrombus formation with secondary fibrin deposition in the femoral head in dysbaric osteonecrosis (7). Li et al. also showed endothelial cell damage as well as a high coagulant and a low fibrinolytic milieu the possible pathologic mechanisms of glucocorticoid-induced ANFH (8). It is noteworthy that in a recent study, alpha-2-

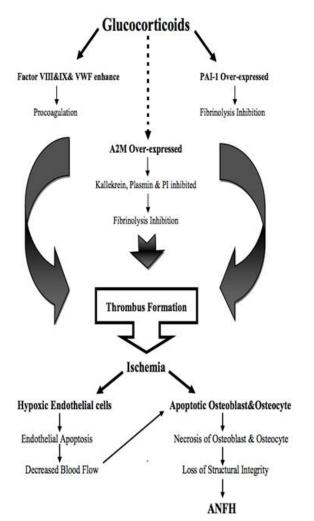


Figure 1. Different mechanisms of glucocorticoid effect on ANFH.

AVASCULAR NECROSIS

macroglobulin (A2M) gene expression was the most significantly upregulated gene in a glucocorticoidinduced ANFH rat model. In this model, glucocorticoids (GCs) seem to modulate significantly the A2M levels (9). GCs can change the A2M gene expression by attaching to a glucocorticoid response element (GRE), a conserved consensus sequence for a putative GC receptor DNA binding site, in the 5'-flanking region proximal to the A2M gene promoter (10). Several other literatures also support the role of GCs in the modulation of A2M (11, 12). A2M has been identified on the luminal surface of endothelial cells in sections of normal human arteries and veins (13). It has also been implicated in hemostasis as a regulator of thrombin and in the development of thromboembolism in children (14-17).

#### Avascular Necrosis and the Coagulation Pathway

Glucocorticoids are the most common non-traumatic cause of ANFH as between 5% to 40% of patients treated with long-term GCs develop ANFH (3, 18). Furthermore, with regards to the regulatory effect of GCs on procoagulation mediators (Factor VIII & IX & VWF) and fibrinolysis inhibitor mediator (PAI-1), there is a possible role of GCs on coagulation (19-21).

Eduardo Ramacciotti et al. in 2010 introduced Alpha-2-Macroglobulin (A2M) as a protein influencing on thrombosis through inflammation, cell shedding, inhibition of fibrinolysis, and hemostatic plug formation. On the other hand, it could play an important role in thrombogenesis and fibrinolysis. It works as an inhibitor of fibrinolysis by inhibiting plasmin and kallikrein, and as an inhibitor of coagulation by inhibiting thrombin (22). Thus, it can be concluded that GCs can make changes in endothelial function both by modulation in A2M gene expression and through thrombosis formation that subsequently induces ischemia [Figure 1].

Björkman et al. in two separate studies in 2004 and 2005 suggested that the coagulation abnormalities in the form of factor V Leiden and prothrombin 20210A gene mutation might play a role in avascular necrosis of the knee and hip (23, 24). Zalavras et al. in 2004 also studied the thrombophilic factor V G1691A mutation (factor V Leiden) and G20210A prothrombin mutation in ANFH patients and introduced factor V Leiden, a genetic risk factor for venous thrombosis. This study and similar studies support the hypothesis that intravascular coagulation is a major pathogenetic mechanism of the disease (25).

#### Ischemia in Avascular Necrosis

Femoral head avascular necrosis can occur due to blood supply disruption, which results in hypoxemic injury to the femoral head. Hypoxia-inducible factor-1  $\alpha$  (HIF-1 $\alpha$ ) is a master regulator of cellular response to hypoxia. It has been reported that there is a coordinated upregulation of HIF-1a and Vascular Endothelial Growth Factor (VEGF) expression under hypoxia in both RNA and protein levels after an ischemic event in the cartilage (26). Furthermore, VEGF expression was reduced by 44% after HIF-1 $\alpha$  knockdown by siRNA transfection in conditional

HIF-1 $\alpha$  knock out mice.VEGF is a well characterized angiogenic factor that is activated by hypoxia. All of these observations demonstrated that upregulation of VEGF in chondrocytes following femoral head ischemia is mediated partially through HIF-1 $\alpha$ . These results are suggesting more vessel formation after inducing ischemia in pig femoral head via VEGF upregulated partially through HIF-1 $\alpha$  (27). These findings introduce VEGF as a factor responsible for neovascularization and repair. Ding et al. in 2013 showed that transplantation of HIF-1 $\alpha$  transgenic bone marrow cells (BMCs) that upregulated VEGF and enhanced both angiogenesis and osteogenesis potentially promotes the repair of necrotic area of GCs-induced ANFH (28).

Considering the results, new therapeutic modalities that can stimulate the repair process and restore normal vascular and growth after ischemic necrosis have been developed (29).

#### Angiogenesis in Avascular Necrosis

In ANFH, interruption of angiogenesis is a pathological process that may lead to impairment of the nutrient supply, cell death, and collapse of the bone. However, the process of angiogenesis in ANFH is not well understood (30).

Recent studies have revealed a strong correlation between ANFH and the expression of VEGF, a major inducer of angiogenesis and more importantly in bone formation and repair (31-35). VEGF regulates bone remodelling by attracting endothelial cells and osteoclasts as well as stimulating osteoblast differentiation (36). Other studies also support augmentation of angiogenesis in bone tissue by VEGF gene transfection, which accelerates bone repairing (37, 38). All of the above besides the similar studies suggest that VEGF has a potential therapeutic effect for avascular necrosis of the bone tissue (39-41).

Yang et al. in 2004, co-transfected bFGF and collagen genes and implanted it in a necrotic femoral head of an animal model and showed that angiogenesis in the bone tissue improved. Also they found that the repair process in the necrotic femoral head accelerated significantly (42).

It has also been shown that, nitric oxide regulates bone turn over through osteoblasts and osteoclasts. Nitric oxide production is impaired by the T-786C eNOS single nucleotide polymorphism. This leads to vasoconstriction, platelet aggregation, reduced angiogenesis, and reduced bone formation, all of which may be associated with avascular necrosis of the hip. In a study by Glueck et al. in 2007, the association of T-786C eNOS polymorphism and resultant reduction of nitric oxide production was shown tocontribute in the pathogenesis of idiopathic ANFH (43).

## Lipid Biosynthesis in Avascular Necrosis

Many studies suggest that several factors are important in the etiology of ANFH, including GCs usage, alcoholism, infections, coagulation defects, and some autoimmune disorders. However, etiological and pathological mechanisms of ANFH have not been yet thoroughly investigated. As mentioned earlier, vascular hypothesis AVASCULAR NECROSIS

is considered to be most persuasive mechanisms among others to date (1). Several studies have identified that hyperlipidemia in the femoral head induced by GCs and alcohol use is associated with ANFH.

Kim et al. in 2008 evaluated the association between sterol regulatory element binding factor (SREBP-2) gene polymorphisms and the susceptibility of ANFH in the Korean population. SREBPs, which belong to the basic helix-loop-helix family of transcription factors, are important in lipogenesis, adipocyte development, and cholesterol homeostasis (44). Also in 2009 Lee et al. reported a polymorphism in intron 7 of the SREBP-1 gene associated with an increased risk of ANFH (45). This would provide the evidence of an association between ANFH and lipid metabolism.

In a study by Okazaki et al. in 2009, It has been shown that GCs-induced ANFH could be caused by disruption of the immune system via LPS-activated toll-like receptor 4 (TLR4) signaling pathway associated with a disruption of the innate immune system and lipid synthesis or metabolism in a rat model (46).

#### **Apoptosis in Avascular Necrosis**

The osteocyte is the most numerous and longestliving cell in bone. They have an active role in regulating bone homeostasis. For example, they can act as an orchestrator of bone remodeling through regulation of osteoblasts and osteoclasts. It has been reported that increased numbers of osteocytes undergo apoptosis in glucocorticoid- and alcohol-induced avascular necrosis (47-49). GCs and alcohol could have direct toxic effects on bone cells, and cause them to become apoptotic (50). The exact mechanisms of non-traumatic avascular necrosis remain elusive; many studies in the past decade have shown that the pathogenesis involves different pathways and factors (51). It was found that the expression of OPG, RANK, and RANKL was higher in the necrotic part than in the normal region in osteonecrotic samples (52). Another study showed different expression levels of bone morphogenetic protein (BMP) in the normal and necrotic sites of femoral heads in patients with avascular necrosis (53). Jun et al. in 2014 reported that the expression level of inducible nitric oxide synthase (iNOS) increased in osteonecrotic samples in compare to control samples suggesting that more nitric oxide was being produced in osteonecrotic samples. They also found that the apoptosis of numerous osteocytes in the avascular necrosis group was mainly associated with the consistently high expression level of iNOS. Furthermore, they supposed that using aminoguanidine, an inhibitor of iNOS, can reduce the production of iNOS in osteocytes and thus cause a reduction in apoptosis of osteocytes. They demonstrated that inhibition of iNOS could prevent non-traumatic avascular necrosis in an ANFH animal model (54). However, the apoptosis signalling pathway is not only mediated through the mitochondrion, but also the extracellular signals can activate caspase through Fas/CD95, and this pathway may play a significant role in the apoptosis of osteocytes in nontraumatic ANFH (55).

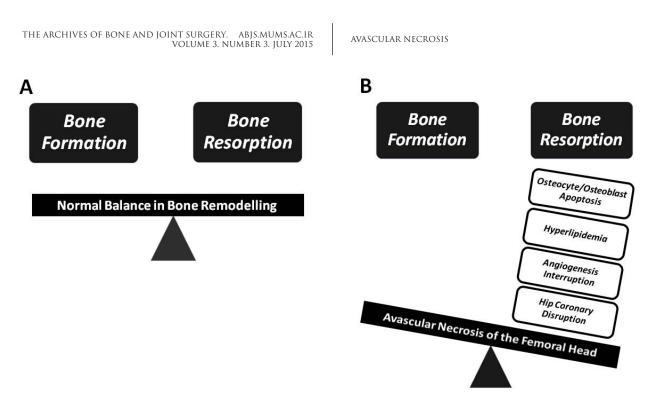


Figure 2. Normal balance between bone formation and bone resorption resulting in bone tissue integrity (A). Bone remodeling balance switch into bone resorption, resulting in loss of femoral head collapse that leads to ANFH (B).

#### **Bone Remodelling in Avascular Necrosis**

Osteoprotegerin (OPG), Receptor Activator of Nuclear Factor kappa-B (RANK) and RANK ligand (RANKL) regulate the balance between osteoclasts and osteoblasts. The expression of these genes affects the maturation and function of osteoblasts and osteoclasts and bone remodelling (56). Samara et al. in 2014 suggested different expression mechanisms for OPG, RANK and RANKL.They could play an important role in the progress of bone remodelling in the necrotic area (52).

Bone morphogenetic proteins (BMPs) are other key proteins in regulating bone remodelling and healing (53). Implantation of osteogenic BMPs such as BMP-2 and BMP-7 at an osseous or extraosseous site results in bone and cartilage formation. These BMPs act primarily as differentiation factors, turning responsive mesenchymal cells into cartilage- and bone-forming cells (57). Implantation of human BMP-2 gene transfected bone marrow stem cells (BMSCs) can repair early-stage experimental femoral head necrosis (41). Thus, these osteogenic BMPs could be used as therapeutic targets for ANFH treatment.

#### Inherited Avascular Necrosis

Although most cases of ANFH are sporadic, some idiopathic familial cases have been reported so far with some families having multiple affected members. Although genetic factors have been implicated in the etiology of ANFH, the causal gene has not been identified yet (58). COL2A1 mutations have been reported to be associated with ANFH particularly in bilateral cases in an autosomal dominant inheritance pathway in a Japanese family. The mutation (p.G1170S) leads to an amino acid change that perturbs a Gly-X-Y triple-helix repeat, which is fundamental structure in type II collagen. It has also been reported that abnormal large-diameter collagen fibrils in the epiphyseal cartilage of ANFH patients was present (59). This implies that abnormal type II collagen could be the cause of inherited ANFH (58). In another study, in 2008, Peiqiang et al. reported the same mutation of COL2A1 responsible for pathology confined to the hip joint, which presents as an isolated precocious hip osteoarthritis, ANFH, and Legg-Calve-Perthes disease in an age at onset dependent manner.

Bone remodelling involves the removal of mineralized bone by osteoclasts followed by the formation of bone matrix by the osteoblasts that subsequently become mineralized. Recent studies have showed that a balance in bone remodelling is maintained in the microenvironment of the femoral head. The regulation of bone remodelling is both local and systemic. The major systemic regulators include growth hormone, glucocorticoids, parathyroid hormone, thvroid hormones, and sex hormones. As far as local regulation of bone remodelling is concerned, many cytokines and growth factors including OPG/RANK/RANKL that affect bone cell functions have been recently identified (60). The balance between bone resorption and bone formation can maintain the integrity of the bone microenvironment. The regulators switch the balance into increasing bone resorption and decreasing bone formation in the bone microenvironment of femoral head, thus, femoral head will collapse [Figure2]. All of

the mechanisms previously described can lead to ANFH by switching the balance into bone resorption.

Further investigations assessing the crossroad of the main molecular pathways are recommended.

# Acknowledgment

The authors are thankful to the Vice Chancellor of Research, Mashhad University of Medical Sciences,

AVASCULAR NECROSIS

Mashhad, Iran, for the financial support.

#### Farzaneh Pouya MD

Mohammad Amin Kerachian MD Medical Genetics Research Center, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran Department of Medical Genetics, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

References

- 1. Kerachian MA, Harvey EJ, Cournoyer D, Chow TY, Seguin C. Avascular necrosis of the femoral head: vascular hypotheses. Endothelium . 2006; 13(4):237-44.
- 2. Weinstein RS, Nicholas RW, Manolagas SC. Apoptosis of osteocytes in glucocorticoid-induced osteonecrosis of the hip. J Clin Endocrinol Metab. 2000; 85(8):2907-12.
- 3. Aldridge JM, 3rd, Urbaniak JR. Avascular necrosis of the femoral head: etiology, pathophysiology, classification, and current treatment guidelines. Am J Orthop (Belle Mead NJ). 2004; 33(7):327-32.
- 4. Assouline-Dayan Y, Chang C, Greenspan A, Shoenfeld Y, Gershwin ME. Pathogenesis and natural history of osteonecrosis. Semin Arthritis Rheum. 2002; 32(2):94-124.
- 5. Vadasz Z, Misselevich I, Norman D, Peled E, Boss JH. Localization of vascular endothelial growth factor during the early reparative phase of the rats' vessels deprivation-induced osteonecrosis of the femoral heads. Exp Mol Pathol. 2004; 77(2):145-8.
- 6. Jacobs B. Epidemiology of traumatic and nontraumatic osteonecrosis. Clin Orthop Relat Res. 1978; (130):51-67.
- Slichter SJ, Stegall P, Smith K, Huang TW, Harker LA. Dysbaric osteonecrosis: a consequence of intravascular bubble formation, endothelial damage, and platelet thrombosis. J Lab Clin Med. 1981; 98(4):568-90.
- 8. Li Y, Chen J, Zhang Z, Wang K, Tong Z, Yan H. The experimental study on treatment of glucocorticoidinduced ischemic necrosis of femoral head by gu fu sheng capsule. J Tradit Chin Med. 2004; 24(4):303-7.
- 9. Kerachian MA, Cournoyer D, Harvey EJ, Chow TY, Begin LR, Nahal A, et al. New insights into the pathogenesis of glucocorticoid-induced avascular necrosis: microarray analysis of gene expression in a rat model. Arthritis Res Ther. 2010;12(3):124.
- 10. Northemann W, Shiels BR, Braciak TA, Hanson RW, Heinrich PC, Fey GH. Structure and acute-phase

regulation of the rat alpha 2-macroglobulin gene. Biochemistry. 1988; 27(26):9194-203.

- 11. Shibahara M, Nishida K, Asahara H, Yoshikawa T, Mitani S, Kondo Y, et al. Increased osteocyte apoptosis during the development of femoral head osteonecrosis in spontaneously hypertensive rats. Acta Med Okayama. 2000; 54(2):67-74.
- 12. Gordon AH, Limaos EA. Effects of bacterial endotoxin and corticosteroids on plasma concentrations of alpha 2 macroglobulin, haptoglobin and fibrinogen in rats. Br J Exp Pathol. 1979;60(4):434-40.
- Becker CG, Harpel PC. alpha2-Macroglobulin on human vascular endothelium. J Exp Med. 1976; 144(1):1-9.
- 14. Cvirn G, Gallistl S, Koestenberger M, Kutschera J, Leschnik B, Muntean W. Alpha 2-macroglobulin enhances prothrombin activation and thrombin potential by inhibiting the anticoagulant protein C/ protein S system in cord and adult plasma. Thromb Res. 2002; 105(5):433-9.
- 15. Ma H, Li R, Zhang Z, Tong T. mRNA level of alpha-2-macroglobulin as an aging biomarker of human fibroblasts in culture. Exp Gerontol. 2004; 39(3):415-21.
- 16. Branson HE, Endo Y, Fagin AR, Schlutz M. Heritable  $\alpha$ 2-Macroglobulin Deficiency in a Patient With Arterial Thrombosis:  $\alpha$ 2-Macroglobulin Deficiency Irvine. J Natl Med Assoc. 1984; 76(11):1107.
- 17. Beheiri A, Langer C, During C, Krumpel A, Thedieck S, Nowak-Gottl U. Role of elevated alpha2-macroglobulin revisited: results of a case-control study in children with symptomatic thromboembolism. J Thromb Haemost. 2007; 5(6):1179-84.
- Koo KH, Kim R, Kim YS, Ahn IO, Cho SH, Song HR, et al. Risk period for developing osteonecrosis of the femoral head in patients on steroid treatment. Clin Rheumatol. 2002; 21(4):299-303.
- 19. van Zaane B, Nur E, Squizzato A, Gerdes VE, Buller HR, Dekkers OM, et al. Systematic review on the effect of glucocorticoid use on procoagulant, anti-coagulant

and fibrinolytic factors. J Thromb Haemost. 2010; 8(11):2483-93.

- 20. Van Zaane B, Nur E, Squizzato A, Dekkers OM, Twickler MT, Fliers E, et al. Hypercoagulable state in Cushing's syndrome: a systematic review. J Clin Endocrinol Metab. 2009;94(8):2743-50.
- 21. Yamamoto Y, Ishizu A, Ikeda H, Otsuka N, Yoshiki T. Dexamethasone increased plasminogen activator inhibitor-1 expression on human umbilical vein endothelial cells: an additive effect to tumor necrosis factor-alpha. Pathobiology. 2004;71(6):295-301.
- 22. Ramacciotti E, Hawley AE, Wrobleski SK, Myers DD Jr, Strahler JR, Andrews PC, et al. Proteomics of microparticles after deep venous thrombosis. Thromb Res. 2010; 125(6): 269-74.
- 23. Bjorkman A, Svensson PJ, Hillarp A, Burtscher IM, Runow A, Benoni G. Factor V leiden and prothrombin gene mutation: risk factors for osteonecrosis of the femoral head in adults. Clin Orthop Relat Res. 2004; (425):168-72.
- 24. Bjorkman A, Burtscher IM, Svensson PJ, Hillarp A, Besjakov J, Benoni G. Factor V Leiden and the prothrombin 20210A gene mutation and osteonecrosis of the knee. Arch Orthop Trauma Surg. 2005; 25(1):51-5.
- 25. Zalavras CG, Vartholomatos G, Dokou E, Malizos KN. Genetic background of osteonecrosis: associated with thrombophilic mutations?. Clin Orthop Relat Res. 2004 (422):251-5.
- 26. Kim HK, Bian H, Aya-ay J, Garces A, Morgan EF, Gilbert SR. Hypoxia and HIF-1alpha expression in the epiphyseal cartilage following ischemic injury to the immature femoral head. Bone. 2009;45(2):280-8.
- 27. Zhang C, Li Y, Cornelia R, Swisher S, Kim H. Regulation of VEGF expression by HIF-1alpha in the femoral head cartilage following ischemia osteonecrosis. Scientific reports. 2012; 2:650.
- 28. Ding H, Gao YS, Hu C, Wang Y, Wang CG, Yin JM, et al. HIF-1alpha transgenic bone marrow cells can promote tissue repair in cases of corticosteroid-induced osteonecrosis of the femoral head in rabbits. PloS one. 2013; 8(5): 63628.
- 29. Kim HK, Bian H, Randall T, Garces A, Gerstenfeld LC, Einhorn TA. Increased VEGF expression in the epiphyseal cartilage after ischemic necrosis of the capital femoral epiphysis. J Bone Miner Res. 2004; 19(12):2041-8.
- 30. Mwale F, Wang H, Johnson AJ, Mont MA, Antoniou J. Abnormal vascular endothelial growth factor expression in mesenchymal stem cells from both osteonecrotic and osteoarthritic hips. Bull NYU Hosp

AVASCULAR NECROSIS

Jt Dis. 2011;69 Suppl 1:S56-61.

- 31. Chim SM, Tickner J, Chow ST, Kuek V, Guo B, Zhang G, et al. Angiogenic factors in bone local environment. Cytokine Growth Factor Rev. 2013;24(3):297-310.
- 32. Vincenzi B, Napolitano A, Zoccoli A, Iuliani M, Pantano F, Papapietro N, et al. Serum VEGF levels as predictive marker of bisphosphonate-related osteonecrosis of the jaw. J Hematol Oncol. 2012; 5:56.
- 33. Li J, Fan L, Yu Z, Dang X, Wang K. The effect of deferoxamine on angiogenesis and bone repair in steroid-induced osteonecrosis of rabbit femoral heads. Exp Biol Med (Maywood). 2014; 240(2):273-80.
- 34. Schipani E, Ryan HE, Didrickson S, Kobayashi T, Knight M, Johnson RS. Hypoxia in cartilage: HIF-1alpha is essential for chondrocyte growth arrest and survival. Genes Dev. 2001; 15(21):2865-76.
- 35. Peng H, Wright V, Usas A, Gearhart B, Shen HC, Cummins J, et al. Synergistic enhancement of bone formation and healing by stem cell-expressed VEGF and bone morphogenetic protein-4. J Clin Invest. 2002; 110(6):751-9.
- 36. Deckers MM, Karperien M, van der Bent C, Yamashita T, Papapoulos SE, Lowik CW. Expression of vascular endothelial growth factors and their receptors during osteoblast differentiation. Endocrinology. 2000;141(5):1667-74.
- 37. Yang C, Yang S, Du J, Li J, Xu W, Xiong Y. Experimental study of vascular endothelial growth factor gene therapy for avascular necrosis of the femoral head. J Huazhong Univ Sci Technolog Med Sci. 2003; 23(3):297-9,
- 38. Yang C, Yang S, Du J, Li J, Xu W, Xiong Y. Vascular endothelial growth factor gene transfection to enhance the repair of avascular necrosis of the femoral head of rabbit. Chin Med J (Engl). 2003; 116(10):1544-8.
- 39. Cao K, Huang W, An H, Jiang DM, Shu Y, Han ZM. Deproteinized bone with VEGF gene transfer to facilitate the repair of early avascular necrosis of femoral head of rabbit. Chin J Traumatol. 2009; 12(5):269-74.
- 40. Zhang GL, Zhang M. Progress of vascularized bone grafting for the treatment of avascular osteonecrosis of the femoral head. Zhongguo Gu Shang. 2008; 21(7):556-8.
- 41. Liu BY, Zhao DW. Treatment for osteonecrosis of femoral head by hVEGF-165 gene modified marrow stromal stem cells under arthroscope. Zhonghua Yi Xue Za Zhi. 2009; 89(37):2629-33.
- 42. Yang C, Yang SH, Du JY, Li J, Xu WH, Xiong YF. Basic

fibroblast growth factor gene transfection to enhance the repair of avascular necrosis of the femoral head. Chin Med Sci J. 2004; 19(2):111-5.

- 43. Glueck CJ, Freiberg RA, Oghene J, Fontaine RN, Wang P. Association between the T-786C eNOS polymorphism and idiopathic osteonecrosis of the head of the femur. J Bone Joint Surg Am. 2007; 89(11):2460-8.
- 44. Kim TH, Baek JI, Hong JM, Choi SJ, Lee HJ, Cho HJ, et al. Significant association of SREBP-2 genetic polymorphisms with avascular necrosis in the Korean population. BMC Med Genet. 2008; 9:94.
- 45. Lee HJ, Choi SJ, Hong JM, Lee WK, Baek JI, Kim SY, et al. Association of a polymorphism in the intron 7 of the SREBF1 gene with osteonecrosis of the femoral head in Koreans. Ann Hum Genet. 2009;73(1):34-41.
- 46. Okazaki S, Nishitani Y, Nagoya S, Kaya M, Yamashita T, Matsumoto H. Femoral head osteonecrosis can be caused by disruption of the systemic immune response via the toll-like receptor 4 signalling pathway. Rheumatology (Oxford). 2009; 48(3):227-32.
- 47. Tsuji M, Ikeda H, Ishizu A, Miyatake Y, Hayase H, Yoshiki T. Altered expression of apoptosis-related genes in osteocytes exposed to high-dose steroid hormones and hypoxic stress. Pathobiology. 2006;73(6):304-9.
- 48. Maurel DB, Boisseau N, Benhamou CL, Jaffre C. Alcohol and bone: review of dose effects and mechanisms. Osteoporos Int. 2012; 23(1):1-16.
- 49. Weinstein RS. Glucocorticoid-induced osteonecrosis. Endocrine. 2012;41(2):183-90.
- 50. Mutijima E, De Maertelaer V, Deprez M, Malaise M, Hauzeur JP. The apoptosis of osteoblasts and osteocytes in femoral head osteonecrosis: its specificity and its distribution. Clin Rheumatol. 2014;33(12):1791-5.
- 51. Fessel J. There are many potential medical therapies

AVASCULAR NECROSIS

for atraumatic osteonecrosis. Rheumatology (Oxford). 2013;52(2):235-41.

- 52. Samara S, Dailiana Z, Chassanidis C, Koromila T, Papatheodorou L, Malizos KN, et al. Expression profile of osteoprotegerin, RANK and RANKL genes in the femoral head of patients with avascular necrosis. Exp Mol Pathol. 2014; 96(1):9-14.
- 53. Samara S, Dailiana Z, Varitimidis S, Chassanidis C, Koromila T, Malizos KN, et al. Bone morphogenetic proteins (BMPs) expression in the femoral heads of patients with avascular necrosis. Mol Biol Rep. 2013; 40(7):4465-72.
- 54. Wang J, Kalhor A, Lu S, Crawford R, Ni JD, Xiao Y. iNOS expression and osteocyte apoptosis in idiopathic, non-traumatic osteonecrosis. Acta Orthop.. 2015; 86(1):134-41.
- 55. Kogianni G, Mann V, Ebetino F, Nuttall M, Nijweide P, Simpson H, et al. Fas/CD95 is associated with glucocorticoid-induced osteocyte apoptosis. Life Sci. 2004; 75(24):2879-95.
- 56. Liu JZ, Ji ZL, Chen SM. [The OPG/RANKL/RANK system and bone resorptive disease]. Sheng Wu Gong Cheng Xue Bao. 2003;19(6):655-60.
- 57. Ebara S, Nakayama K. Mechanism for the action of bone morphogenetic proteins and regulation of their activity. Spine (Phila Pa 1976). 2002; 27(16 Suppl 1):10-5.
- 58. Miyamoto Y, Matsuda T, Kitoh H, Haga N, Ohashi H, Nishimura G, et al. A recurrent mutation in type II collagen gene causes Legg-Calve-Perthes disease in a Japanese family. Hum Genet. 2007; 121(5):625-9.
- 59. Ponseti IV, Maynard JA, Weinstein SL, Ippolito EG, Pous JG. Legg-Calve-Perthes disease. Histochemical and ultrastructural observations of the epiphyseal cartilage and physis. J Bone Joint Surg Am. 1983; 65(6):797-807.
- 60. Hadjidakis DJ, Androulakis ll. Bone remodeling. Ann N Y Acad Sci. 2006;1092(1):385-96.