

**CASE REPORT**

# Contribution of Sonication to the Identification of Megaprosthesis Infection in Culture Negative Oncologic Patient

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*Research performed at British Hospital of Buenos Aires, Argentina**Received: 24 December 2022 Accepted: 12 January 2023***Abstract**

Conventional central osteosarcoma mainly affects the metaphysis of long bones in young people. The use of megaprotheses in oncological patients has increased in recent years. However, this type of surgery is not exempt from complications, with infections being the most common. In recent years, the presence of biofilm-forming bacteria has increased. Biofilm characteristics allow bacteria to resist hostile environmental conditions. The application of long wave ultrasound (process known as sonication) on the rescued inert material before culture interrupts the biofilm and generates a significantly higher recovery of bacterial growth compared to conventional tissue culture. We present the case of a 12-year-old patient with osteosarcoma of the femur, who, after surgery, developed a prosthetic infection detected by sonication, with negative soft tissue culture.

**Level of evidence:** IV**Keywords:** Megaprotheses, Osteosarcoma, Prosthetic infection, Sonication**Introduction**

Conventional central osteosarcoma is the most common primary malignant bone tumor in children, adolescents, and young adults,<sup>1</sup> arising mainly in the metaphysis of the long bones and gradually invading the epiphysis and diaphysis and eventually the entire joint space.<sup>2</sup> The most frequently affected sites are the distal femur and the proximal tibia. With increasing survival rates after chemotherapy, limb-sparing surgeries are now the standard of care for most of these osteosarcomas.<sup>3,4</sup> Due to the great bone loss after tumor resection, especially in those areas near a joint, prosthetic reconstruction is required in most patients. This concept is mainly applied in those skeletally immature patients, with the aim of preserving joint function as much as possible.<sup>5</sup> The use of modular megaprotheses in cancer patients has gradually increased.<sup>6</sup> Since its inception in the 80s, they have been used for the reconstruction of bone defects after tumor resection thanks to their immediate fixation and ability to allow weight bearing.<sup>7</sup> On the other hand, this type of implant is not without complications; one of the most frequent is infection, present in 3% to 30% of cases.<sup>9,10</sup>

To apply the right antimicrobial treatment, pathogen identification is essential. Most common techniques rely on

the recovery of samples from periprosthetic tissue followed by inoculation in culture media.<sup>10</sup> However, this technique may be interfered with by certain factors that decrease its sensitivity, such as the negligent use of previous antibiotics, sampling errors, inadequate amounts of bacteria or inaccurate transport.<sup>11</sup> Another reason for the failure of microbial culture is the presence of bacteria organized in biofilms. This concept refers to certain complex communities of microorganisms that can be found attached to a surface. Biofilm characteristics allow bacteria to resist hostile environmental conditions. For these reasons, they are considered a major cause of persistent nosocomial infections in immunocompromised patients; and can prevent microbiological diagnosis and eradication of the microorganism due to poor response to antibiotic treatment.<sup>12, 13</sup> The presence of this biofilm on implant surfaces is responsible for the preservation of implant-related infections, since it has the ability to alter the activation of the phagocytic and complement immune system.<sup>14, 15</sup>

In this sense, the application of long wave ultrasound (process known as sonication) on the rescued inert material (implant, plastic, prosthesis) before culture interrupts the

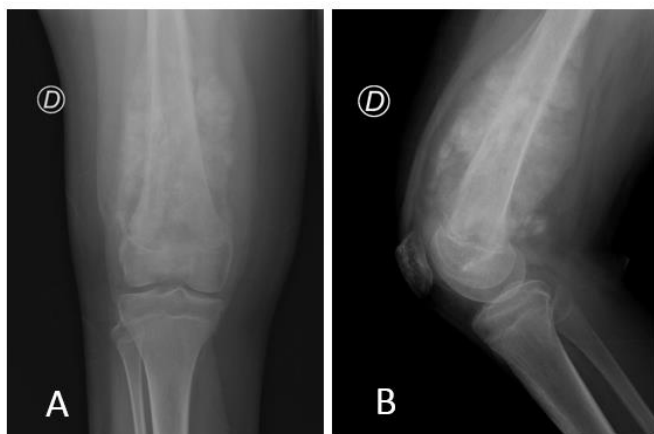
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biofilm and generates a significantly higher recovery of bacterial growth compared to conventional tissue culture.<sup>16</sup> We present the case of an oncological patient with postoperative pain, with subsequent biopsy without microbiological recovery, and later positive culture after sonication of the extracted material, confirming periprosthetic infection.

### Case Presentation

We present the case of a 12-year-old female patient with a clinical history of grade I obesity (BMI: 32). She consulted our service in July 2020 due to atraumatic right gonalgia of 10 days of evolution, which gradually increased in intensity, with gait intolerance at the time of consultation. He presented with an increase in the diameter of the right thigh at distal level with knee flexion up to 50° limited by pain. X-rays of the right knee were requested showing a heterogeneous radiopaque image involving the lateral cortex of the diaphysis, metaphysis, physis and epiphysis of the right femur, with soft tissue involvement in the medial, lateral, anterior and posterior region of the distal third and partially in the middle third of the right femur [Figure 1 A-B]. With a strong suspicion of a diagnosis of osteosarcoma, in the first instance, metastatic lesions were ruled out.



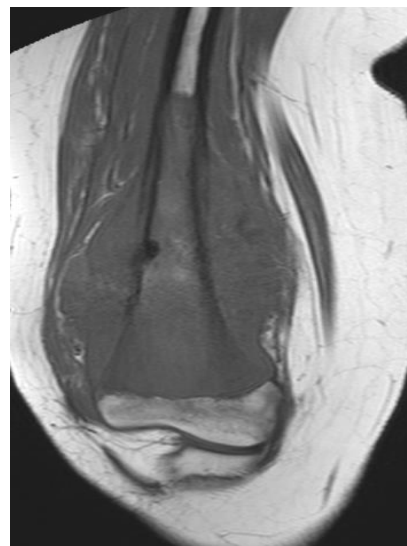
**Figure 1.** X-rays of the right knee showing a heterogeneous radiopaque image involving the lateral cortex of the diaphysis, metaphysis, physis and epiphysis of the right femur, with soft tissue involvement in the medial, lateral, anterior and posterior region of the distal third and partially in the middle third of the right femur

A nuclear magnetic resonance of the right thigh and knee was requested, which reported a bone lesion of pathological aspect. It presented a heterogeneous signal, with endomedullary involvement, significant periosteal reaction and associated soft tissue component [Figure 2].

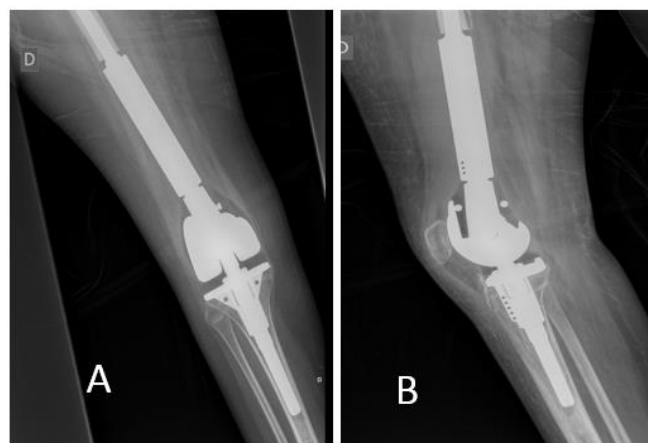
A bone puncture was performed under CT and the histopathological substrate confirmed the diagnosis of conventional central osteosarcoma.

Fifteen days after the diagnosis of osteosarcoma, the patient receives the first infusion of doxorubicin - cisplatin as neoadjuvant therapy. Two days later, she intercurrent with coronavirus infection in August 2020. The patient did not receive neoadjuvant therapy for 25 days because at that time, the interaction of the coronavirus with chemotherapy drugs was unknown. After that period, she received the

second dose of chemotherapy infusion; however, 10 days later, she had another coronavirus infection. In view of this situation, after an interdisciplinary meeting, it was decided to perform oncologic resection surgery with free margins and reconstruction with megaprosthesis in October 2020 [Figure 3 A-B].



**Figure 2.** MNR (STIR) of the right thigh and knee which reported a bone lesion of pathological aspect. It presented a heterogeneous signal, with endomedullary involvement, significant periosteal reaction and associated soft tissue component



**Figure 3.** X-ray of right knee after oncologic resection surgery and reconstruction with megaprosthesis

After surgery, and during adjuvant therapy, the patient had numerous hospital admissions due to fever, mucositis and adverse effects of medication, in addition to two coronavirus infections.

Eight months after surgery, she referred us to right gonalgia secondary to physical exertion that gradually increased in intensity. On physical examination he presented with non-

deficient palpatory pain, with no clinical signs of infection. A laboratory was requested for infection parameters with white blood cell count of 3900 mm<sup>3</sup>, C-reactive protein of 0.6 mg/dL and erythrocyte sedimentation rate of 7 mm/h. Pain was continuously increasing, with no improvement with NSAIDs or opioids. Control X-rays were requested, showing alteration in the structure compatible with signs of acute periostitis around the femoral prosthetic component [Figure 4]. Due to persistent pain, a bone and soft tissue biopsy was performed, without microbiological recovery. In view of this picture of increased pain and radiological signs with the presence of periostitis, the prosthesis was converted to a right knee spacer due to suspicion of chronic prosthetic infection [Figure 5 A-B]. In this surgery, culture samples were sent to the microbiology service and the prosthetic plastic component was sent for culture by sonication. The patient underwent intravenous antibiotic treatment initially with clindamycin / ceftazidime for a week and then started oral antibiotic therapy. Twelve days after the intervention, the soft tissue cultures sent were still negative, but the material sent by sonication obtained positive culture results for *Staphylococcus epidermidis* and *Corynebacterium aurimucosum*. The patient underwent prophylactic antibiotic treatment for 4 months and after this period the spacer was converted to a right knee megaprosthesis [Figure 6 A-D]. Currently, the patient has been 18 months since the last intervention. She is walking without the use of assistance, with infectious laboratory parameters within normal values, without reporting any pain.

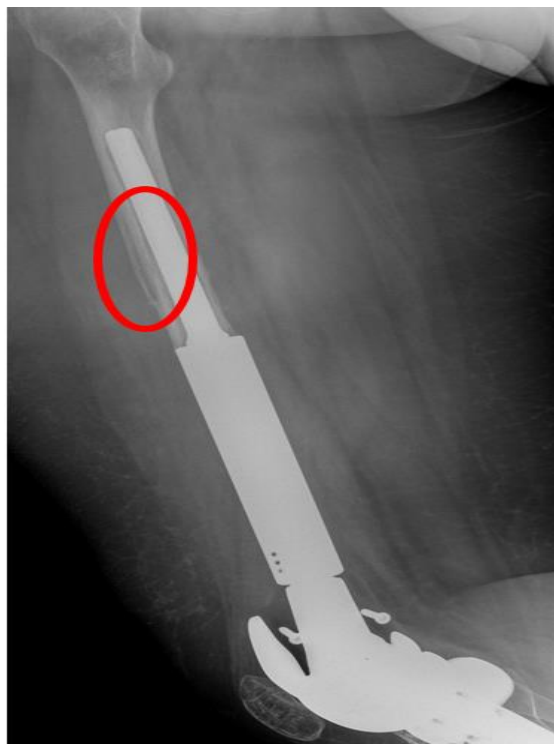


Figure 4. X-ray showing alteration in the structure compatible with signs of acute periostitis around the femoral prosthetic component (red circle)

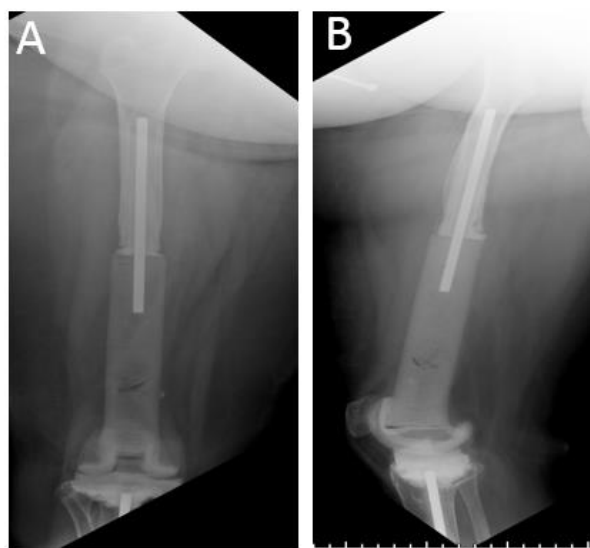


Figure 5. X-ray of right knee spacer



Figure 6. X-ray of right knee megaprosthesis

## Discussion

Periprosthetic infections are one of the most frequent complications after joint replacement, especially in modular megaprotheses; with rates from 3% to more than 30%.<sup>17</sup> considering this, the detection of the infectious microorganism is absolutely necessary to choose the appropriate antibiotics and improve the pathogen elimination route.

Currently, the usual conventional method of diagnosis is the culture of tissue and periprosthetic fluid.<sup>18</sup> However, the presence of biofilms, prior antibiotic exposure and the effect of previous surgery may render standard cultures falsely negative.<sup>18</sup> The use of gentle ultra-sonication to disrupt adherent microbes thanks to biofilms formed was first used on peritoneal catheters<sup>19</sup> and ureteral stents.<sup>20</sup> Tunney et al.<sup>21</sup> were the first to use this method in prosthetic implants as a complementary method in the diagnosis of infection. In the study carried out by Trampuz et al.<sup>22</sup> on a total of 79 patients diagnosed with periprosthetic infection, 62 cases (78.48%) had positive cultures by the traditional method and by sonication; while the remaining 17 cases (21.52%) only had positive cultures by sonication method. In another

article published by Ribeiro et al.,<sup>23</sup> out of a total number of 148 prostheses analyzed from patients with suspected prosthetic infection, 67% had positive culture results by sonication method, while only 39% did so by traditional culture methods. In our case, cultures by traditional method were negative, while the prosthetic component analyzed by sonication was positive for *Staphylococcus epidermidis* and *Corynebacterium aurimucosum*. Purchner et al.<sup>24</sup> evaluated whether sonication culture improved the microbiological diagnosis of periprosthetic megaprosthesis infections compared to conventional tissue culture. Out of a total of 31 megaprotheses analyzed by both methods, in 25 (80.64%) pathogens were detected by sonication and 12 (38.70%) were detected by tissue culture. *Staphylococcus epidermidis* was the most frequently detected microorganism in both groups (n: 11 in the sonication group and n: 5 in the tissue group). On the other hand, there are few published studies on *Corynebacterium* and periprosthetic infections, most of which were performed before the identification of recently described *Corynebacterium* species. In 2004, Roux published a study of *Corynebacterium* species isolated from bone and joint infections.<sup>25</sup> Of the 31 patients reported, 8 had joint prosthesis infection (2 each with *C. amycolatum* and *C. striatum*, 3 with *C. aurimucosum*, and 1 with *C. jeikeium*).

### Conclusion

In conclusion, bacterial culture by sonication method is nowadays one of the most sensitive methods in the diagnosis of joint prosthesis infections.

The negative culture of preoperative joint aspiration and periprosthetic soft tissues obtained intraoperatively does not exclude the presence of bacteria in the implants. Where possible, orthopedic surgeons should use this method to confirm or refute the diagnosis of periprosthetic infection, especially in cases of suspected chronic low-grade infection.

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### References

- Ottaviani G, Jaffe N. The epidemiology of osteosarcoma. *Cancer Treat Res.* 2009; 152:3-13. doi: 10.1007/978-1-4419-0284-9\_1.
- Chen Y, Yu XC, Xu SF, et al. Impacts of tumor location, nature and bone destruction of extremity osteosarcoma on selection of limb salvage operative procedure. *Orthop Surg.* 2016; 8(2): 139-149. doi: 10.1111/os.12237.
- Hegyi M, Semsei AF, Jakab Z, et al. Good prognosis of localized osteosarcoma in young patients treated with limb-salvage surgery and chemotherapy. *Pediatr Blood Cancer.* 2011; 57(3): 415-422. doi: 10.1002/pbc.23172.
- Gharedaghi M, Peivandi MT, Mazloomi M, Shoorin HR, Hasani M, Seyf P, Khazaei F. Evaluation of Clinical Results and Complications of Structural Allograft Reconstruction after Bone Tumor Surgery. *Arch Bone Jt Surg.* 2016 Jun;4(3):236-42. PMID: 27517069; PMCID: PMC4969370.
- Eleutério SJ, Senerchia AA, Almeida MT, et al. Osteosarcoma in patients younger than 12 years old without metastases have similar prognosis as adolescent and young adults. *Pediatr Blood Cancer.* 2015; 62(7): 1209-1213. doi: 10.1002/pbc.25459.
- Ercolano LB, Christensen T, McGough R, et al. Treatment solutions are unclear for perimegaprosthetic infections. *Clin Orthop Relat Res.* 2013; 471(10):3204-3213. doi: 10.1007/s11999-013-2852-7.
- Ahlmann ER, Menendez LR, Kermani C, et al. Survivorship and clinical outcome of modular endoprosthetic reconstruction for neoplastic disease of the lower limb. *J Bone Joint Surg Br.* 2006; 88(6):790-795. doi: 10.1302/0301-620X.88B6.17519.
- Capanna R, Morris HG, Campanacci D, et al. Modular uncemented prosthetic reconstruction after resection of tumours of the distal femur. *J Bone Joint Surg Br.* 1994; 76(2):178- 186. PMID: 8113272.
- Gosheger G, Goetze C, Harges J, et al. The influence of the alloy of megaprotheses on infection rate. *J Arthroplasty.* 2008; 23(6):916-920. doi: 10.1016/j.arth.2007.06.015.
- Shahpari O, Mousavian A, Elahpour N, Malahias MA, Ebrahimzadeh MH, Moradi A. The Use of Antibiotic Impregnated Cement Spacers in the Treatment of Infected Total Joint Replacement: Challenges and Achievements. *Arch Bone Jt Surg.* 2020 Jan;8(1):11-20. doi: 10.22038/abjs.2019.42018.2141. PMID: 32090140; PMCID: PMC7007713.
- Padgett DE, Silverman A, Sachjowicz F, et al. Efficacy of intraoperative cultures obtained during revision total hip arthroplasty. *J Arthroplasty.* 1995; 10(4):420-6. doi: 10.1016/s0883-5403(05)80140-8.
- Davies D. Understanding biofilm resistance to antibacterial agents. *Nat Rev Drug Discov.* 2003; 2(2):114-22. doi: 10.1038/nrd1008.
- Singh PK, Schaefer AL, Parsek MR, et al. Quorum-sensing signals indicate that cystic fibrosis lungs are infected with bacterial biofilms. *Nature.* 2000; 407(6805):762-4. doi: 10.1038/35037627.
- Portillo ME, Corvec S, Borens O, et al. *Propionibacterium acnes*: an underestimated pathogen in implant-associated infections. *BioMed Res Int.* 2013; 2013:804391. doi:

- 10.1155/2013/804391.
15. Lass R, Giurea A, Kubista B, et al. Bacterial adherence to different components of total hip prosthesis in patients with prosthetic joint infection. *Int Orthop*. 2014; 38(8):1597-1602. doi: 10.1007/s00264-014-2358-2.
  16. Carmen JC, Roeder BL, Nelson JL, et al. Treatment of biofilm infections on implants with low-frequency ultrasound and antibiotics. *Am J Infect Control*. 2005; 33(2):78-82. doi: 10.1016/j.ajic.2004.08.002.
  17. Sevelde F, Schuh R, Hofstaetter JG, et al. Total femur replacement after tumor resection: limb salvage usually achieved but complications and failures are common. *Clin Orthop Relat Res*. 2015; 473(6):2079-2087. doi: 10.1007/s11999-015-4282-1.
  19. Taha M, Abdelbary H, Ross FP, et al. New Innovations in the Treatment of PJI and Biofilms - Clinical and Preclinical Topics. *Curr Rev Musculoskelet Med*. 2018; 11(3):380-388. doi: 10.1007/s12178-018-9500-5.
  20. Gorman SP, Adair CG, Mawhinney WM. Incidence and nature of peritoneal catheter biofilm determined by electron and confocal laser scanning microscopy. *Epidemiol Infect*. 1994; 112(3):551-9. doi: 10.1017/s0950268800051256.
  21. Keane PF, Bonner MC, Johnston SR, et al. Characterization of

- biofilm and encrustation on ureteric stents in vivo. *Br J Urol*. 1994; 73(6):687-91. doi: 10.1111/j.1464-410x.1994.tb07557.x.
22. Tunney MM, Patrick S, Gorman SP, et al. Improved detection of infection in hip replacements. A currently underestimated problem. *J Bone Joint Surg Br*. 1998; 80(4):568-72. doi: 10.1302/0301-620x.80b4.8473.
  23. Trampuz A, Piper KE, Jacobson MJ, et al. Sonication of removed hip and knee prostheses for diagnosis of infection. *N Engl J Med*. 2007; 357(7):654-63. doi: 10.1056/NEJMoa061588.
  24. Ribeiro TC, Honda EK, Daniachi D, et al. The impact of sonication cultures when the diagnosis of prosthetic joint infection is inconclusive. *PLoS One*. 2021; 16(7):e0252322. doi: 10.1371/journal.pone.0252322.
  25. Puchner SE, Döring K, Staats K, et al. Sonication culture improves microbiological diagnosis of modular megaprotheses. *J Orthop Res*. 2017; 35(7):1383-1387. doi: 10.1002/jor.23406.
  26. Roux V, Drancourt M, Stein A, et al. *Corynebacterium* species isolated from bone and joint infections identified by 16S rRNA gene sequence analysis. *J. Clin. Microbiol*. 2004; 42(5):2231-2233. doi: 10.1128/JCM.42.5.2231-2233.2004.